1. INTENDED USE

Bovine Albumin ELISA Kit is a sandwich ELISA system for quantitative measurement of bovine albumin. This is intended for research use only.

2. STORAGE AND EXPIRATION

When the complete kit is stored at 2-8°C, the kit is stable until the expiration date shown on the label on the box. Opened reagents should be used as soon as possible to avoid loss in optimal assay performance caused by storage environment.

3. INTRODUCTION

Albumin is mostly a simple hydrophilic protein present in cells and body fluids. Albumin is synthesized in the liver, and serum albumin (69 kDa, pl 4.9) occupies 56-60% of total serum proteins. Because of its large population, albumin is very important in maintaining plasma osmotic pressure. Albumin can bind hydrophobic physiological substances e.g. fatty acids, bilirubin, and thyroxine and contributes the transfer of these substances. Bovine albumin is generally thought to be an inert protein and is commonly used in various incubation media, and especially calf serum is very important component of tissue culture media. This means that if some substances are prepared using tissue- or cell-culture technique, they possible contaminated with bovine albumin. In order to assure product purity bovine albumin in the product has to be estimated. The present highly sensitive ELISA kit will be useful in this kind of estimation.
4. ASSAY PRINCIPLE

In Shibayagi’s bovine Albumin ELISA Kit, standards or samples are incubated in monoclonal anti-albumin antibody-coated wells to capture albumin. After 1 hour incubation and washing, HRP (horse radish peroxidase)-conjugated anti-albumin antibody is added, and incubated for 1 hour. After washing, HRP-complex remaining in wells is reacted with a chromogenic substrate (TMB) for 30 minutes, and reaction is stopped by addition of acidic solution, and absorbance of yellow product is measured spectrophotometrically at 450 nm. The absorbance is nearly proportional to albumin concentration. The standard curve is prepared by plotting absorbance against standard albumin concentrations. Albumin concentrations in unknown samples are determined using this standard curve.

5. PRECAUTIONS

- For professional use only. Beginners are advised to use this kit under the guidance of experienced person.
- Do not drink, eat or smoke in the areas where assays are carried out.
- This kit contains components of animal origin. These materials should be handled as potentially infectious.
- Be careful not to allow the reagent solutions of the kit to touch the skin, eyes and mucus membranes. Especially be careful for the reaction stopper because it is 1 M sulfuric acid. The reaction stopper and the substrate solution may cause skin/eyes irritation. In case of contact with these wash skin/eyes thoroughly with water and seek medical attention, when necessary.
- Avoid contact with the acidic Reaction stopper solution and Chromogenic substrate reagent containing hydrogen peroxide and tetramethylbenzidine (TMB). Wear gloves and eye and clothing protection when handling these reagents. The materials must not be pipetted by mouth.
- Unused samples and used tips should be rinsed in 1% formalin, 2% glutaldehyde, or more than 0.1% sodium hypochlorite solution for more than 1 hour, or be treated by an autoclave before disposal.
- Dispose consumable materials and unused contents in accordance with applicable regional/national regulatory requirements.
- Use clean laboratory glassware.
- In order to avoid dryness of wells, contamination of foreign substances and evaporation of dispensed reagents, never forget to cover the well plate with a plate cover supplied, during incubation.
- ELISA can be easily affected by your laboratory environment. Room temperature should be at 20-25°C strictly. Avoid airstream velocity over 0.4 m/sec. (including wind from air conditioner) (*①), and humidity less than 30%.
- For more details, watch our web movie [Assay circumstance].

6. REAGENTS SUPPLIED

<table>
<thead>
<tr>
<th>Components</th>
<th>State</th>
<th>Amount</th>
</tr>
</thead>
<tbody>
<tr>
<td>A. Anti-albumin-coated plate</td>
<td>Ready for use.</td>
<td>96 wells/1 plate</td>
</tr>
<tr>
<td>B. Standard bovine albumin (500 ng/ml) (derived from bovine)</td>
<td>Concentrated. Use after dilution</td>
<td>200 µl/1 vial</td>
</tr>
<tr>
<td>C. Buffer solution</td>
<td>Ready for use.</td>
<td>60 ml/1 bottle</td>
</tr>
<tr>
<td>D. HRP-conjugated anti-albumin antibody</td>
<td>Concentrated. Use after dilution.</td>
<td>200 µl/1 vial</td>
</tr>
<tr>
<td>E. Chromogenic substrate reagent (TMB)</td>
<td>Ready for use.</td>
<td>12 ml/1 bottle</td>
</tr>
<tr>
<td>F. Reaction stopper (1M H₂SO₄)</td>
<td>Ready for use.</td>
<td>12 ml/1 bottle</td>
</tr>
<tr>
<td>I. Concentrated washing buffer (10x)</td>
<td>Concentrated Use after dilution.</td>
<td>100 ml/1 bottle</td>
</tr>
<tr>
<td>Plate cover</td>
<td>-</td>
<td>1 plate</td>
</tr>
<tr>
<td>Instruction Manual</td>
<td>-</td>
<td>1 copy</td>
</tr>
</tbody>
</table>

7. EQUIPMENTS OR SUPPLIES REQUIRED BUT NOT SUPPLIED

- Use as a check box
- Purified water (distilled water)
- Test tubes for preparation of standard solution series.
- Glassware for dilution of washing buffer (a graduated cylinder, a bottle)
- Pipettes (disposable tip type). One should be able to deliver 50 µl precisely, and another for 50-500 µl.
- Syringe-type repeating dispenser like Eppendorf multipette plus which can dispense 100 µl.
- Paper towel to remove washing buffer remaining in wells.
- A vortex-type mixer.
- A shaker for 96 well-plate (600-1200 rpm)
• An automatic washer for 96 well-plate (if available), or a wash bottle with a jet nozzle (refer to our web movie [Washing of microplate]).
• A 96 well-plate reader (450 nm ±10 nm, 620 nm: 600-650 nm)
• Software for data analysis, if available. Shibayagi is proposing the use of assay results calculation template for EXCEL. Please check our website (http://www.shibayagi.co.jp/en/tech_003.html).

8. PREPARATION OF REAGENTS

✧ Bring all reagents of the kit to room temperature (20-25°C) before use.
✧ Prepare reagent solutions in appropriate volume for your assay. Do not store the diluted reagents.

Concentrated reagents
Standard bovine albumin (500 ng/ml)
Below is an example of preparing each standard solution.

<table>
<thead>
<tr>
<th>Volume of standard solution</th>
<th>Buffer solution</th>
<th>Concentration (ng/ml)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Original solution: 50 μl</td>
<td>450 μl</td>
<td>50.0</td>
</tr>
<tr>
<td>50 ng/ml solution: 250 μl</td>
<td>250 μl</td>
<td>25.0</td>
</tr>
<tr>
<td>25 ng/ml solution: 250 μl</td>
<td>250 μl</td>
<td>12.5</td>
</tr>
<tr>
<td>12.5 ng/ml solution: 250 μl</td>
<td>250 μl</td>
<td>6.25</td>
</tr>
<tr>
<td>6.25 ng/ml solution: 250 μl</td>
<td>250 μl</td>
<td>3.13</td>
</tr>
<tr>
<td>3.13 ng/ml solution: 250 μl</td>
<td>250 μl</td>
<td>1.56</td>
</tr>
<tr>
<td>1.56 ng/ml solution: 250 μl</td>
<td>250 μl</td>
<td>0.78</td>
</tr>
<tr>
<td>0 (Blank)</td>
<td>250 μl</td>
<td>0</td>
</tr>
</tbody>
</table>

HRP-conjugated anti-albumin antibody
Prepare working solution by dilution of (D) with the buffer solution (C) to 1:100.

Concentrated washing buffer (10x)
Dilute 1 volume of the concentrated washing buffer (10x) to 10 volume with deionized water to prepare working solution. Example: 100 ml of concentrated washing buffer (10x) and 900ml of dionized water.
Storage and stability

Anti-albumin-coated plate
If seal is not removed, put the strip back in a plastic bag with zip-seal originally used for well-plate container and store at 2-8°C. The strip will be stable until expiration date.

Standard bovine albumin (500 ng/ml)
Standard solutions prepared should be used as soon as possible, and should not be stored.
Dispose remaining prepared solution.

Buffer solution and Chromogenic substrate reagent
Use only volume you need for your assay. Remaining reagents should be stored at 2-8°C fastening the cap tightly. It remains stable until expiration date. Once opened, we recommend using as soon as possible to avoid influence by environmental condition.

HRP-conjugated anti-albumin antibody
Unused working solution (already diluted) should be disposed. The rest of the undiluted solution: if stored tightly closed at 2-8°C, it is stable until expiration date.

Reaction stopper (1 M H$_2$SO$_4$)
Close the stopper tightly and store at 2-8°C. It maintains stability until expiration date.

Concentrated washing buffer (10x)
The rest of undiluted buffer: if stored tightly closed at 2-8°C, it is stable until expiration date.
Dispose any unused diluted buffer.

9. TECHNICAL TIPS

- In manual operation, proficiency in pipetting technique is recommended.
- The reagents are prepared to give accurate results only when used in combination within the same box. Therefore, do not combine the reagents from kits with different lot numbers. Even if the lot number is the same, it is best not to mix the reagents with those that have been preserved for some period.
• Be careful to avoid any contamination of assay samples and reagents. We recommend the use of disposal pipette tips, and 1 tip for 1 well.
• Optimally, the reagent solutions of the kit should be used immediately after reconstitution. Otherwise, store them in a dark place at 2-8°C.
• Time the reaction from the pipetting of the reagent to the first well.
• Prepare a standard curve for each assay.

• Dilution of the assay sample must be carried out using the buffer solution provided in the kit.
• The chromogenic substrate reagent (TMB) should be almost colorless before use. It turns blue during reaction, and gives yellowish color after addition of reaction stopper. Greenish color means incomplete mixing.
• To avoid denaturation of the coated antibody, do not let the plate go dry.
• As the antibody-coated plate is module type of 8 wells x 12 strips, each strip can be separated by cutting the cover sheet with a knife and used independently.
• When ELISA has to be done under the airstream velocity of over 0.4 m/sec. and the humidity of less than 30%, completely close each well in addition to cover the well plate with a plate cover in each step of incubation.
• Ex.) Cover the well plate with parafilm, and put the plate cover on it. Or place the well plate with the plate cover in an incubator, or in a styrofoam box. Take the best way depending on situation of each laboratory. For more details, watch our web movie [Assay circumstance].

10. PREPARATION OF SAMPLES

This kit is intended to measure minute albumin in bovine samples such as tissue- or cell-culture products. Adjust your samples’ pH within 6.5-8.0. A small sample volume 100 μl is needed as standard procedure.

Note that this kit is too sensitive to measure bovine serum or plasma. If you would like to measure them, they should be diluted about 1000000x. They need to be diluted using the kit’s buffer so as to be within the assay range, 0.78-50 ng/ml.

Samples should be immediately assayed or stored below −35°C until assay. Before starting assay, stir thawed samples sufficiently. Do not repeat freeze-and-thaw cycles. Dilution of a sample should be made in a test tube using buffer solution prior to adding them to wells. Turbid samples or those containing insoluble materials should be centrifuged before testing to remove any particulate matter.
Testing for compatibility of your samples with Shibayagi’s kit using a positive sample.

Due to various factors of your sampling conditions (anesthesia, preservatives, anticoagulants, raised sample pH caused by loss of CO₂ during standing and storage, preservative used, evaporation and condensation during storage in a freezer, etc), sometimes the kit does not work well with your samples. If the standard curve is in a good shape, while your samples give low absorbance, please check the compatibility of your samples (serum or plasma) by a simple recovery test as follows.

Place 90 μl of your sample (e.g. a sample from control group in your experiment) in a small test tube, then add 10 μl of the highest standard solution. Assay this mixture together with the original sample, and compare the assay values. The assay value of the mixture will be around [0.9 x original sample + 0.1 x highest standard concentration]. If the assay value is increased as expected, the assay system is working well with your sample.

Especially when you use Shibayagi’s kit for the first time, we recommend you to run this simple recovery test.

11. ASSAY PROCEDURE

Remove the cover sheet of the antibody-coated plate after bringing up to room temperature.

1. Wash the anti-albumin-coated plate (A) by filling the wells with washing buffer and discard 4 times(*②), then strike the plate upside-down onto folded several sheets of paper towel to remove residual buffer in the wells.
2. Pipette 100 μl of standard solution to the wells designated for standards, and 100 μl of samples to the designated sample wells.
3. Shake the plate gently on a plate shaker(*③).
4. Put a plate cover on the plate and incubate for 1 hour at 20-25°C.
5. Discard the reaction mixture and rinse wells as step (1).
6. Pipette 100 μl of HRP-conjugated anti-albumin antibody to all wells, and shake as step (3).
7. Put a plate cover on the plate and incubate the plate for 1 hour at 20-25°C.
8. Discard the reaction mixture and rinse wells as step (1).
9. Pipette 100μl of Chromogenic substrate reagent to wells, and shake as step (3).
10. Put a plate cover on the plate and incubate the plate for 30 minutes at 20-25°C.
11. Add 100 μl of the reaction stopper to all wells to stop the coloration.
12. Measure the absorbance of each well at 450 nm (reference wavelength, 620*nm) using a plate reader within 30 minutes.
*Refer to the page 7 for notes of *② and *③.

12. CALCULATIONS

1. Prepare a standard curve by plotting standard concentration on X-axis and absorbance on Y-axis. (Refer to our web site for more detailed explanation about standard curve. Shibayagi is offering a convenient Excel template. http://www.shibayagi.co.jp/en/tech_003.html)

2. Using the standard curve, read the albumin concentration of a sample at its absorbance*, and multiply the assay value by dilution factor if the sample has been diluted. Though the assay range is wide enough, in case the absorbance of some samples is higher than that of the highest standard, please repeat the assay after proper dilution of samples with the buffer solution.
* We recommend the use of 3rd order regression curve for log-log plot, or 4 parameters method for log-normal plot in computer calculation.

Physiological or pathological situation of bovine should be judged comprehensively taking other examination results into consideration.

Bovine Albumin assay standard curve (an example)
Absorbance may change due to assay environment.
13. PERFORMANCE CHARACTERISTICS

- Assay range
  The assay range of the kit is 0.78 ~ 50 ng/ml.

- Specificity
  The antibodies used in this kit are specific to albumin.

<table>
<thead>
<tr>
<th>Sample</th>
<th>Cross reaction</th>
</tr>
</thead>
<tbody>
<tr>
<td>Bovine albumin</td>
<td>100%</td>
</tr>
<tr>
<td>Mouse albumin</td>
<td>Less than 0.05%</td>
</tr>
<tr>
<td>Rat albumin</td>
<td>Less than 0.05%</td>
</tr>
<tr>
<td>Human albumin</td>
<td>Less than 0.05%</td>
</tr>
</tbody>
</table>

- Precision of assay
  Within assay variation (4 samples, 5 replicates assay), Mean CV is less than 5%

- Reproducibility
  Between assay variation (3 samples, 4 days, 3 replicates assay), Mean CV is less than 5%

- Recovery test
  Standard albumin was added in 3 concentrations to serum sample and assayed. The recoveries were 91 ~104%

- Dilution test
  Serum sample was serially diluted by 4 steps. The dilution curves showed linearity with R² = 0.9977 ~ 0.9996.

14. TROUBLE SHOOTING

- Low absorbance in all wells
  Possible explanations:
  1. The standard or samples might not be added.
  2. Reagents necessary for coloration such as HRP-conjugated anti-albumin antibody or Chromogenic substrate reagent might not be added.
  3. Wrong reagents related to coloration might have been added. Wrong dilution of HRP-conjugated anti-albumin antibody.
  4. Contamination of enzyme inhibitor(s).
5. Influence of the temperature under which the kits had been stored.
6. Excessive hard washing of the well plate.
7. Addition of chromogenic substrate reagent soon after taking out from a refrigerator might cause poor coloration owing to low temperature.
   - Blank OD was higher that that of the lowest standard concentration (0.78 ng/ml).

Possible explanations:
Improper or inadequate washing. (Change washing frequency from 4 times to 5-8 times at the constant stroke after the reaction with HRP-conjugated anti-albumin antibody.)

- High coefficient of variation (CV)
Possible explanation:
1) Improper or inadequate washing.
2) Improper mixing of standard or samples.
3) Pipetting at irregular intervals.

- Q-1: Can I divide the plate to use it for the other testing?
  A-1: Yes, cut off the clear seal on the plate with cutter along strip. Put the residual plate, which is still the seal on, in a refrigerator soon

- Q-2: I found 96 well-plate is empty when I opened the box.
  A-2: As this kit is dried type, not preservation stabilizer is added.
  For detailed FAQs and explanations, refer to “Trouble shooting and Important Points in Shibayagi’s ELISA kits” on our website (http://www.shibayagi.co.jp/en/tech_004.html).

Summary of assay procedure: Use as a check box
*First, read this instruction manual carefully and start your assay after confirmation of details.
For more details, watch our web movie [ELISA by MOVIE] on our website.
Bring the well-plate and all reagents back to 20-25°C for 2 hours.
Concentrated washing buffer must be diluted to 10 times by purified water that returned to 20-25°C.
Standard solution dilution example:

<table>
<thead>
<tr>
<th>Concentration (ng/ml)</th>
<th>50</th>
<th>25</th>
<th>12.5</th>
<th>6.25</th>
<th>3.13</th>
<th>1.56</th>
<th>0.78</th>
<th>0</th>
</tr>
</thead>
<tbody>
<tr>
<td>Standard solution (µl)</td>
<td>Orig. sol. 50</td>
<td>250*</td>
<td>250*</td>
<td>250*</td>
<td>250*</td>
<td>250*</td>
<td>250*</td>
<td>250*</td>
</tr>
<tr>
<td>Buffer solution (µl)</td>
<td>450</td>
<td>250</td>
<td>250</td>
<td>250</td>
<td>250</td>
<td>250</td>
<td>250</td>
<td>250</td>
</tr>
</tbody>
</table>

*One rank higher standard.

Preparation of the positive sample and samples.
<table>
<thead>
<tr>
<th>Step</th>
<th>Description</th>
</tr>
</thead>
<tbody>
<tr>
<td>↓Washing 4 times*②)</td>
<td>Anti-albumin-coated plate (Dried-plate)</td>
</tr>
<tr>
<td>Samples, or Standards</td>
<td>100 μl</td>
</tr>
<tr>
<td>↓Shaking (*③), Incubation for 1 hours at 20-25°C. (Standing (*④))</td>
<td>Dilute HRP-conjugated anti-albumin antibody (D) to 100x with buffer (C) returned to 20-25°C.</td>
</tr>
<tr>
<td>↓Washing 4 times*</td>
<td></td>
</tr>
<tr>
<td>HRP-conjugated anti-albumin antibody 100 μl</td>
<td></td>
</tr>
<tr>
<td>↓Shaking (*③) Incubation for 1 hours at 20-25°C. (Standing (*④))</td>
<td>Dilute reagents during the first reaction.</td>
</tr>
<tr>
<td>↓Washing 4 times* (*②)</td>
<td></td>
</tr>
<tr>
<td>Chromogenic substrate reagent (TMB) 100 μl</td>
<td>After dispense, the color turns to blue depending on the concentration.</td>
</tr>
<tr>
<td>↓Shaking(*③), Incubation for 30 minutes at 20-25°C. (Standing (*④))</td>
<td>After dispense, the color turns to yellow depending on the concentration.</td>
</tr>
<tr>
<td>Reaction stopper ( 1M H₂SO₄) 100 μl</td>
<td>Immediately shake.</td>
</tr>
<tr>
<td>↓Shaking(*③)</td>
<td></td>
</tr>
<tr>
<td>Measurement of absorbance (450 nm, Ref 620 nm) *⑤</td>
<td>Ref. wave cancels the dirt in the back of plate.</td>
</tr>
</tbody>
</table>

*② Guideline of washing volume: 300 μl/well for an automatic washer and for a pipette if the washing buffer is added by pipette. In case of washing by using 8 channel pipette, sometimes the background tends to be high. If so, change washing frequency from 4 times to 5-8 times at the constant stroke after the reaction with HRP conjugated streptavidin.

Standard of plate-washing pressure: 5-25 ml/min. (Adjust it depending on the nozzle’s diameter.) Refer to our web movie [Washing of microplate].

*③ Guideline of shaking: 600-1,200 rpm for 10 seconds x 3 times.

*④ Put a plate cover on the plate during the reaction after shaking.
*⑤ 600-650 nm can be used as reference wavelength.
*⑥ After removal of wash buffer, immediately dispense the next reagent.
*⑦ Refer to our web movie [Handling of pipetting].
*⑧ Refer to our web movie [Assay circumstance].

Worksheet example

<table>
<thead>
<tr>
<th>Strip 1&amp;2</th>
<th>Strip 3&amp;4</th>
<th>Strip 5&amp;6</th>
<th>Strip 7&amp;8</th>
<th>Strip 9&amp;10</th>
<th>Strip 11&amp;12</th>
</tr>
</thead>
<tbody>
<tr>
<td>A</td>
<td>Pos. Control</td>
<td>Sample 8</td>
<td>Sample 16</td>
<td>Sample 24</td>
<td>Sample 32</td>
</tr>
<tr>
<td>B</td>
<td>Sample 1</td>
<td>Sample 9</td>
<td>Sample 17</td>
<td>Sample 25</td>
<td>Sample 33</td>
</tr>
<tr>
<td>C</td>
<td>Sample 2</td>
<td>Sample 10</td>
<td>Sample 18</td>
<td>Sample 26</td>
<td>Sample 34</td>
</tr>
<tr>
<td>D</td>
<td>Sample 3</td>
<td>Sample 11</td>
<td>Sample 19</td>
<td>Sample 27</td>
<td>Sample 35</td>
</tr>
<tr>
<td>E</td>
<td>Sample 4</td>
<td>Sample 12</td>
<td>Sample 20</td>
<td>Sample 28</td>
<td>Sample 36</td>
</tr>
<tr>
<td>F</td>
<td>Sample 5</td>
<td>Sample 13</td>
<td>Sample 21</td>
<td>Sample 29</td>
<td>Sample 37</td>
</tr>
<tr>
<td>G</td>
<td>Sample 6</td>
<td>Sample 14</td>
<td>Sample 22</td>
<td>Sample 30</td>
<td>Sample 38</td>
</tr>
<tr>
<td>H</td>
<td>Sample 7</td>
<td>Sample 15</td>
<td>Sample 23</td>
<td>Sample 31</td>
<td>Sample 39</td>
</tr>
</tbody>
</table>

Assay worksheet

<table>
<thead>
<tr>
<th>1</th>
<th>2</th>
<th>3</th>
<th>4</th>
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<th>8</th>
<th>9</th>
<th>10</th>
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<tbody>
<tr>
<td>A</td>
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</tbody>
</table>

Storage condition
Store the kit at 2-8°C (Do not freeze).

Term of validity
6 months from production (Expiration date is indicated on the container.)